

Coconut Exosomes from Fructification

C778181

Storage at -80°C (12 months). Avoid freeze/thaw cycle.

Introduction:

1. Coconut is a common tropical fruit, rich in fatty acids, proteins and trace elements. Studies have reported that the active ingredients of coconut have activities such as lowering cholesterol, protecting the heart, and anti-cancer. In addition, coconut extract is also widely used in the field of beauty and cosmetics because of its good inhibitory effect on metalloproteinases. Exosomes derived from coconuts have the advantages of small size and easy availability, and contain active substances such as RNA, fatty acids, and proteins, so they have the potential to be developed into natural preparations.
2. This product is exosomes obtained through separation and purification from coconut extract. It can be used in studies such as experimental control, direct loading and targeted modification, and tissue repair.

Product Specifications:

Name	Coconut Exosomes from Fructification
Specifications	1E+10 Particles/EA
Appearance	White Powder
pH	7.0
Aseptic Testing	Aseptic
Total Protein Concentration	(BCA)
Purity	≥90%

Product Advantages:

1. Strict quality control standards: Multiple quality control analyses are conducted, with stringent standards for various aspects including concentration and purity.
2. Comprehensive characterization results: Exosome raw material products are characterized in three dimensions in accordance with the MISEV2018 guidelines, namely electron microscopic morphology, particle size and particle distribution, and Western Blot (WB) with three positive and one negative protein markers.
3. Wide range of application scenarios: It can be used as a control in experimental procedures and functional experiments; it is applicable to engineering modifications such as targeted modification and molecular loading, as well as the development of exosome therapeutic products.

Case Presentation:

1. NTA detection of exosome particle size distribution and particle concentration:
Shake and disperse the obtained exosomes evenly, dilute them to an appropriate multiple with PBS, mark the sample name and dilution multiple, and complete the dilution preparation of the sample; before testing the sample, test the diluent first: absorb 200 μ L of diluent with a pipette for on-machine detection, and confirm that the components and instruments are in normal operation; after the measurement of the diluent is completed and the test result is normal, start testing the sample, take 200 μ L of the diluted sample for on-machine detection; stop the test when the number of counted particles reaches more than 100, export the data results, and complete the sample detection.
2. Observation of exosome morphology by TEM:
Resuspend exosomes in 50-100 μ L of 2% PFA. Add 5 μ L of the mixed suspension to a Formvar carbon-coated copper grid; alternatively, drop 5-10 μ L of the mixed suspension onto a piece of parafilm and place the copper grid with the Formvar film facing down on the suspension. Prepare 2-3 copper grids for each sample. Add 100 μ L of PBS onto the parafilm. Use tweezers to place the copper grid (with the Formvar film facing down) on the PBS droplets for washing (during all steps, keep the Formvar film surface moist while the other surface remains dry). Place the copper grid on 50 μ L of 1% glutaraldehyde droplets for 5 minutes. Wash the copper grid on 100 μ L of ddH₂O 8 times, 2 minutes each time. Place the copper grid on 50 μ L of uranyl oxalate droplets (pH 7.0) for 5 minutes. Place the copper grid on 50 μ L of methyl cellulose droplets for 10 minutes, operating on ice. Put the copper grid on the stainless steel ring at the top of the sample stage and blot off excess liquid with filter paper. Air-dry for 5-10 minutes. Place the copper grid in the sample box and take electron microscope photos at 80kV.
3. Western Blot Detection of Exosome Markers (Three Positive and One Negative):
Add the isolated and purified exosomes to the lysis buffer (E778170). After lysis, aspirate the supernatant and dilute the sample to an appropriate concentration according to the measured protein concentration. Add 4 $\times$ LDS loading buffer (T466588) to the lysis buffer, boil at 95°C for 5 minutes, and perform a quick centrifugation after cooling to room temperature. Loading: Load all 15 μ L of the sample into the lane, and add markers at the start and end of the sample lanes. Electrophoresis: 190V for 70 minutes. Activate the PVDF membrane with methanol 10 minutes in advance. Membrane transfer: The transfer buffer needs to be pre-cooled in advance, 275mA for 70 minutes. Blocking: Prepare 1% BSA in TBST solution and block at room temperature for 1 hour. Primary antibody incubation: Incubate overnight at 4°C on a shaker; dilute the antibody with 1% BSA in TBST solution. Membrane washing: Wash three times with 1 $\times$ TBST, 10 minutes each time. Secondary antibody incubation: Dilute the antibody with 1% BSA in TBST solution, and incubate on a shaker at room temperature for 1 hour. Membrane washing: Wash three times with 1 $\times$ TBST, 10 minutes each time. Exposure and photography.



Precautions and Disclaimer:

This product is limited to scientific research use by professional personnel. It must not be used for clinical diagnosis or treatment, nor for food or drugs.

